Discovery and Characterization of Hydroxysteroid 17-β Dehydrogenase 13 Inhibitors Anand Balakrishnan, Archie Reyes, Jonathan Lloyd, Manuel Roqueta-Rivera, Mary Chau, Kelsey Garlick, Yat Sun Or and Bryan Goodwin

Enanta Pharmaceuticals, Inc., 500 Arsenal Street, Watertown, MA 02472, USA.

BACKGROUND

The human HSD17B13 gene encodes a 300 amino acid protein annotated as an NAD⁺-dependent 17-β hydroxysteroid dehydrogenase 13 (HSD17β13) with broad substrate specificity. Single nucleotide polymorphisms in the gene lead to reduction in full-length HSD17B13 mRNA and protein levels, and confer a strong protective effect on liver injury, inflammation, fibrosis, and cirrhosis in NASH patients. Consequently, HSD17B13 inhibitors may be beneficial in treating NASH. Here, we present the discovery of the lead series of HSD17B13 inhibitors and the elucidation of their molecular mechanisms of action using enzymology and structural biology.



Figure 1. (A) NAD-dependent HSD17β13-catalyzed reaction, and (B) Crystal structure of dimeric HSD17 β 13 with mobile loops in blue

METHODS

- Materials. Recombinant (His)₆-ENLYFQSG- HSD17β13 was expressed in Sf9 insect cells using a baculoviral expression system and purified using metal affinity purification and size exclusion chromatography. NAD-Glo assay kit was purchased from Promega. HSD17 β 13 compounds were synthesized at Enanta.
- Discovery of lead compounds against HSD17β13. High-throughput screening to identify HSD17β13 inhibitors was performed using acoustic mass spectrometry. Hits were confirmed and characterized using coupled-enzyme luminescence assay to detect NADH and mass spectrometry assays to detect oxidized products.
- **Biochemical Assays.** Activity assays were done in 96w or 384w plates under discontinuous modes of detection: (1) chemiluminescence from the NAD-Glo assay; and (2) product detection via RapidFire mass spectrometry. Assay mixtures contained 40 mM Tris (pH 7.4), 0.01% BSA, 0.01% Tween 20, 50-100 nM enzyme, 10 - 50 µM substrates (LTB4/estradiol), and 0-100 µM compounds.
- **Structural Analysis.** Structure determination using x-ray crystallography was performed with HSD17 β 13 co-crystallized in the presence of inhibitors and/or NAD+.

RESULTS

Discovery of HSD17β13 Inhibitors

• From 502 HTS hits, we have identified 4 lead series that were reconfirmed for their potencies and HSD selectivity via luminescence-NADH assay and RF/MS analysis. Further structural analyses led to Series 1c, 2b and 2c compounds.



The Liver Meeting 2022, AASLD, November 4-8, 2022, Walter E. Washington Convention Center, Washington DC



Lead optimization: Series 1c, 2b, 2c

edions near th

active site

Figure 4. (A) Crystal structure of dimeric NAD-bound HSD17β13, and (B) Zoomed-in substrate binding site of HSD17β13 showing the NAD-binding loop and the mobile C-terminus (both in blue)

RESULTS

D Potent and selective inhibition of HSD17β13 series 1 and 2

Table 2. Potency and selectivity of HSD17β13 Series 1c, 2b and 2c compounds. ¹RF/MS= Rapid Fire Mass Spectrometry with Leukotriene B4; ²RF/MS= RapidFire Mass Spectrometry with E2=Estradiol; ³NADH Luminescence Assay; ⁴Direct measurement of NADH consumption at 340 nm

Assay		1c	2b	2c
Biochemical Activity ^{1,2} Inhibition of product formation (RF/MS)	Human ¹ , IC ₅₀	14 nM	79 nM	37 nM
	Mouse ² , IC ₅₀	2.5 nM	74 nM	9 nM
Cellular Activity² Inhibition of product formation (RF/MS)	HEK293 Human, IC ₅₀	47 nM	34 nM	28 nM
	HEK293 Mouse, IC ₅₀	55 nM	1083 nM	45 nM
HSD Selectivity ³ IC ₅₀ ratio (HSD17βx/ HSD17β13)	HSD17B1	>7000×	>1265×	1838×
	HSD17B2	4357×	322×	322×
	HSD17B4	>7000×	982×	1260×
	HSD17B3, B5, B10, B11	>7000×	>1265×	>2700×
	HSD11B1	>7000×	>630×	>1350×
DH Selectivity IC ₅₀ ratio (Metabolic DH/ HSD17β13)	Human isocitrate dehydrogenase 1 ³	>7000×	>1265×	>2700×
	Rabbit glycerol-3-P dehydrogogenase ⁴	4285x	150×	170×
	Rabbit lactate dehydrogenase ⁴	>7000×	>1265×	190×

 \Box HSD17 β 13 series 1c, 2b and 2c are tight-binding inhibitors

Figure 5. (A) Dependence of IC₅₀ on HSD17 β 13 concentration; and (B) Morrison Fit analysis for tight binding inhibition. IC₅₀ = half-maximal inhibitory concentration, K_i^{app} = apparent inhibition constant. Data are mean ± standard deviation of duplicates.

RESULTS

Table 3. HSD17 β 13 inhibition modality by Series 1c, 2b and 2c. IC₅₀ = half-maximal inhibitory concentration, K_i = inhibition constant and α = inhibitor affinity index. Data are mean ± standard deviation of triplicates.

	Varied NAD+		Varied LTB4			
	<i>K</i> _i (nM)	α	Model	<i>K</i> i (nM)	α	Model
1c	5 ± 0.2	0.7 ± 0.2	Non-competitive	1.7 ± 0.1	11 ± 2	Non-competitive
2 b	135 ± 26	0.7 ± 0.1	Non-competitive	25 ± 14	13 ± 2	Non-competitive
2c	9 ± 3	1.3 ± 0.4	Non-competitive	7.2 ± 0.1	2.2 ± 0.2	Non-competitive

\Box Lead optimized compounds binding at the HSD17 β 13 active site induces conformational changes in the dynamic regions

Figure 9. Series 2 compounds 1c (A) and 2b (B) induce conformational changes in HSD17β13. NADbinding loop and the mobile C-terminus are shown in blue. Residues involved in NAD interactions: V221, T223 and F225. Residues involved in inhibitor interactions: S230, T231 and W234.

CONCLUSION

- Biochemical characterization of HSD17B13 and the development of specific inhibitors will enable a deeper understanding of the pathophysiological function of this enzyme in chronic liver disease.
- We have identified HSD17B13-specific inhibitors and utilized these compounds to probe the enzymology and structural biology of HSD17B13.
- Crystallographic structures of HSD17B13 in complex with substrates/inhibitors shed light on the catalytic function and inhibition of this enzyme.

ACKNOWLEDGEMENTS

We thank Pure Honey Technologies (Billerica, MA) for RF/MS analysis, and Proteros GmBH (Martinsried, Germany) for Structural Biology support.

© 2022 Enanta Pharmaceuticals, Inc.